3,5-Disubstituted-4-hydroxyphenyls Linked to 3-Hydroxy-2-methyl-4(1*H*)-pyridinone: Potent Inhibitors of Lipid Peroxidation and Cell Toxicity

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Received November 3, 1999

Introduction. For many years radical scavenging antioxidants have been successfully used to protect synthetic materials and food products from the degrading process of oxidation. More recently a role as neuroprotective agents for the treatment of disorders known to involve oxidative stress (e.g. stroke, traumatic brain injury, Parkinson's disease, and Alzheimer's disease) has been proposed and supported by animal models. The effectiveness of radical scavengers in reducing oxidative stress within a living biological environment is undermined, however, by the continual production of radicals via mechanisms such as that described by the Fenton reaction, a process that is catalyzed by the presence of Fe²⁺ (eq 1). We postulated

that treatment with both a radical scavenger and an Fe chelator might protect living tissue from oxidative stress to a greater degree than is achievable with either a radical scavenger or an Fe chelator alone (for examples of each, see Chart 1).⁴

A further rationale, that radical scavengers and Fe chelators may be able to interact synergistically,5 is supported by reports of tert-butylphenolic antioxidants interacting with methoxyphenol,6 phosphites,7 ascorbate,8 thiols,8 and sulfites9 to produce synergistic antioxidant effects. In addition it was hypothesized that a significant advantage might be gained over the combined administration of separate radical scavengers and Fe chelators, by combining the structural features for radical scavenging and Fe chelation within one "hybrid" molecule. First, a hybrid molecule could have the potential to behave synergistically via intramolecular interaction, a more favorable process than one which relies upon intermolecular interaction. 10 Second, by creating a hybrid molecule, it was likely that the antioxidant component of the molecule (typically lipophilic in nature, e.g. BHT (1)) would bestow more

Chart 1. Structures of Typical Antioxidant (1, 8, 9) and Fe Chelator (2, 7) Molecules

lipophilicity to the Fe chelator component (typically hydrophilic in nature, e.g. desferrioxamine (2)), giving the molecule a greater potential to penetrate and sequester Fe from areas susceptible to oxidative damage (i.e. lipids, proteins, and DNA).^{11,12}

Chemistry. Favorable structural components for the hybrid molecules were selected by the evaluation of several classes of Fe chelator and radical scavenger in in vitro lipid peroxidation and cell toxicity assays, 13,14 This investigation, together with an assessment of amenability to synthetic manipulation, led to the identification of 3-hydroxy-4(1H)-pyridinone^{15,16} and 2.6disubstituted phenol¹⁷ as preferred structural units for incorporation into hybrid molecules and to the design of target structures 3a-f, where the two structural features are linked via simple alkyl or alkylthio chains. The key intermediate amines **4a**-**f** in the synthesis of 3a-f were prepared in 15-63% overall yield from the commercially available phenols 5a-d, using standard synthetic methods (Scheme 1). The amines 4a-f were then reacted with benzyl maltol (6), 18 followed by deprotection, to give the target compounds 3a-f in 17-56% yield from **4a**-**f** (Scheme 2).

Results and Discussion. Lipid peroxidation in rat brain homogenates was used to measure the antioxidant capacity of the molecules in a biological environment (Table 1).13 All the compounds (3a-f) were more potent inhibitors of lipid peroxidation than the Fe chelator deferiprone (7)¹⁹ or the antioxidants BHT (1), Trolox (8), and LY231617 (9).20 The two 2,6-di-tert-butyl-substituted compounds 3a,f were the most potent inhibitors of lipid peroxidation within the series (3a-f). Replacement of the tert-butyl groups in compounds 3a,b with methoxy groups led to compounds 3d,e and a consequent reduction in potency. An increase in the linker chain length as seen in compounds 3a-c also resulted in a decrease in potency. Substitution of the benzylic CH2 group in 3b with a sulfur atom gave the more potent compound 3f.

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Scheme 1. Synthesis of Amines $4a-f^a$

"Reagents and conditions: i. BH $_3$ -SMe $_2$. THF, reflux, 63%; ii. H $_2$. Pd/C. EtOH, 50 psi, rt. 98%; iii. (a) SOCl $_2$. DMF (cat.), PhMe, CH $_2$ Cl $_2$. rt. (b) NH $_4$ OH, THF, 0 °C, 98%; iv. LiAlH $_4$. Et $_2$ O, reflux, 65%; v. LiAlH $_4$. THF, reflux, 100%; vi. MsCl. Et $_3$ N. CH $_2$ Cl $_2$. rt. 74%; vii. NaCN, DMF, 100 °C, 63%; viii. BH $_3$ -SMe $_2$. THF, reflux, 80%; ix. BnCl, K $_2$ CO $_3$, DMF, 80 °C, 81%; x. MeNO $_2$, NH $_4$ OAc, reflux, 76%; xi. LiAlH $_4$. THF, reflux, 86%; xii. MeCN, KOH, reflux, 46%; xiii. H $_2$. Pd/C, MeOH, rt. 94%; xiv. LiAlH $_4$. THF, reflux, 44%; xv. n-BuLi, TMSCl. THF, -78 °C to rt, 97%; xvi. (a) n-BuLi, S $_8$. THF, -78 to -30 °C, then 2-chloroacetamide. (b) Bu $_4$ NF, MeOH, reflux, 57%; xvii. BH $_3$ -SMe $_2$, THF, reflux, 100%.

Scheme 2. Synthesis of Compounds 3a-f^a

 a Reagents and conditions: i. 5 N NaOH, H₂O, EtOH, **6**, 18 reflux, 24–94%; ii. H₂, Pd/C, EtOH, rt, 18–100% for **3a–e**; iii. BCl₃·SMe₂, CH₂Cl₂, rt, 95% for **3f**.

The ability of the compounds to protect cerebellar granule cells (CGC) from iodoacetate (IAA)-induced toxicity was measured (Table 1). ¹⁴ The 2.6-di-*tert*-butyl-substituted compounds **3a**—**c.f** protected cells from IAA-induced toxicity at lower concentrations than deferiprone (**7**). BHT (**1**), Trolox (**8**), and LY231617 (**9**). The 2,6-dimethoxy-substituted compounds **3d**,**e** were less

Table 1. Biological Activities of Compounds

compd	inhibition of lipid peroxidation ^a IC ₅₀ , µM	protection of CGC from IAA-induced oxidative stress ^b EC ₅₀ , μΜ (rel efficacy) ^c
1	5.9	6.0 (0.9)
$3a^d$	0.3	0.3 (0.9)
$3b^c$	1.0	0.3 (0.8)
3c	2.9	0.6 (0.7)
3d	3.3	33.3 (0.9)
3e	2.0	26.8 (0.8)
$3f^r$	0.4	0.4 (0.6)
7	3.9	46.7 (1.0)
8	28.7	77.8 (0.9)
9	14.8	5.0 (0.9)

"Compounds were tested in duplicate, and results are the average of at least two independent experiments. ^b Compounds were tested in duplicate, and results are the average of at least three independent experiments. ^cRel efficacy: an indication of the percent of viable cells at the maximal efficacious concentration (i.e. 1.0 = 100% viability). ^d Tested as the mesylate salt. ^c Tested as the mesylate salt. Anal. ($C_{23}H_{33}NO_3 \cdot CH_3SO_3H$) C, H: N: calcd, 2.99; found, 2.46. ^c Tested as the hydrochloride salt. Anal. ($C_{22}H_{31}NO_3S \cdot HCl$) C, N; H: calcd, 7.57; found, 7.07.

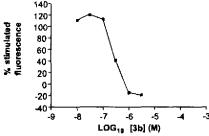


Figure 1. Inhibition of IAA-induced oxidative stress by compound **3b.** The intracellular oxidative stress induced during the IAA cell toxicity assay was measured using the oxidant-sensitive fluorescent dye 2',7'-dichlorodihydrofluorescein diacetate (DCFH-DA). The nonfluorescent DCFH-DA readily crosses cell membranes whereupon it is trapped within the cytoplasm by deacetylation as the non-membrane-permeable form 2',7'-dichlorodihydrofluorescein (DCFH). Upon oxidation, DCFH yields the highly fluorescent product 2',7'-dichlorofluorescein (DCF). Compound **3b** was tested as its mesylate salt and inhibited IAA-induced oxidative stress with an EC₅₀ = 0.28 μ M.

effective than BHT (1) and LY231617 (9) but more effective than deferiprone (7) and Trolox (8). The relative efficacy of compounds $3\mathbf{a} - \mathbf{e}$ decreased with increasing chain length. Within the 2,6-di-*tert*-butyl-substituted series, $3\mathbf{a}$, b provided the best protection against IAA-induced cellular toxicity, in terms of potency and relative efficacy. Using the oxidant-sensitive fluorescent dye 2'.7'-dichlorodihydrofluorescein diacetate (DCFH-DA), the neuronal toxicity induced by IAA was shown to be a result of oxidative stress, and $3\mathbf{b}$ inhibited the oxidation of DCFH to DCF in a similar concentration-dependent manner to its inhibition of IAA-induced cell death, confirming that $3\mathbf{b}$ protects neuronal cells from oxidative stress (Figure 1).²¹

In addition to the significant neuroprotection offered by the 2,6-di-*tert*-butyl compounds 3a-f, compound 3b showed marked enhancement in neuroprotection over the combination of the 3-hydroxy-2-methyl-4(1*H*)-pyridinone 7 and di-*tert*-butylphenol 9. (Note: 9 was chosen as an appropriate comparative agent for 3b because of their similar lipophilicity.) Thus, compound 3b (mlog P=3.9) showed significantly enhanced neuroprotection

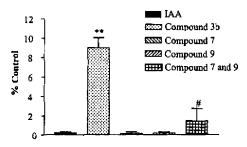


Figure 2. Inhibition of IAA-induced cell toxicity. CGC were exposed to 30 μ M IAA for 30 min in a physiological salt solution. This was replaced with maintenance media containing 1 μ M test compound, and the cells were tested for viability 24 h later. Compound **3b** was tested as its mesylate salt. The error bars represent SE bars. Statistical analysis was performed using two-tailed paired ℓ -test. Statistical significance is defined as p < 0.05; **significantly different from 7 and 9 (p < 0.01); *significantly different from IAA alone (p < 0.05).

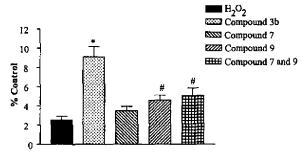


Figure 3. Inhibition of H_2O_2 -induced cell toxicity. CGC were exposed to $100~\mu M$ H_2O_2 for 10 min prior to addition of test compounds at a concentration of $10~\mu M$. The cells were tested for viability 24 h later. Compound 3b was tested as its mesylate salt. The error bars represent SE bars. Statistical analysis was performed using two-tailed paired t-test. Statistical significance is defined as p < 0.05; *significantly different from T and T0 (p < 0.05); *significantly different from T1.

over the dual administration of **7** (mlog P=-0.16) and **9** (mlog P=3.8) in two models of chemical-induced cell toxicity: the IAA cell toxicity assay¹⁴ (Figure 2) and a H_2O_2 cell toxicity assay²² (Figure 3).

Conclusion. A greater understanding of the complex multicomponent processes and mechanisms underlying neurodegenerative disorders such as stroke, traumatic brain injury, Parkinson's disease, and Alzheimer's disease has encouraged a growing trend to produce neuroprotective drugs with more than one mechanism of action.²³ This Communication describes the covalent linking of 3,5-disubstituted-4-hydroxyphenyls with 3-hydroxy-2-methyl-4(1 H)-pyridinone, to produce molecules that are potent inhibitors of lipid peroxidation and cell toxicity. Compound 3b (CEB-1370)24 achieved its neuroprotective effect via inhibition of oxidative stress and displayed a superior neuroprotective action compared to the dual administration of the radical scavenger, ditert-butylphenol 9,20 and the Fe chelator, 1.2-dimethyl-3-hydroxy-4(1*H*)-pyridinone 7.19 Compounds of this series are currently under evaluation for the treatment of neurodegenerative disorders, and further data will be published in due course.25

Acknowledgment. We thank Ken Heatherington for his thorough analytical chemistry support and Ian A. Cliffe for his help in preparing this manuscript.

Supporting Information Available: Chemistry and biology experimental details are available free of charge via the Internet at http://pubs.acs.org.

References

- (i) (a) Branen, A. L.; Davidson, P. M. Use of Antioxidants in Self-preserving Cosmetic and Drug Formulations. Cosmet. Sci. Technol. Ser. 1997, 16, 159-179. (b) Sims, R. J.; Fioritti, J. A. Antioxidants. Gen. Foods (USA), Biotechnol. Food Ingredients 1991, 483-505.
- (2) Marciniak, G.: Petty, M. A. Design and Biological Evaluation of New Antioxidants for use in Cerebrovascular Disorders. *Drugs Future* 1996, 21, 1037–1046.
- (3) Graf, E.; Mahoriey, J. R.; Bryant, R. G.; Eaton, J. W. Iron-catalyzed Hydroxyl Radical Formation. J. Biol. Chem. 1984, 259, 3820–3624
- (4) This postulate was made despite a report that no additional protection is seen in a cardiac ischemia model when a combination of the Fe chelator deferiprone and the phenolic antioxidant (+)-cyanidanol-3 was administered. See: van der Kraaij, A. M. M.; van Eijk, H. G.; Koster, J. F. Prevention of Postischemic Cardiac Injury by the Orally Active Iron Chelator 1.2-Dimethyl 3-Hydroxy-4-Pyridone (L.1) and the antioxidant (+)-Cyanidanol 3. Circulation 1989, 80, 158-164.
- (5) After our work, a report demonstrating intermolecular synergy between ferriheme-bound drugs and chain-breaking antioxidants, in an erythrocyte membrane peroxidation assay, has confirmed our rationale. See: Dailly, E.; Urien, S.; Tillement, J. P. Chain-Breaking Antioxidants and Ferriheme-Bound Drugs are Synergistic Inhibitors of Erythrocyte Membrane Peroxidation. Free Radical Res. 1998, 28, 205-214.
- (6) Mahoney, L. R.; DaRooge, M. A. Inhibition of Free-Radical Reactions. IV. The Synergistic Effect of 2,6-Di-tert-butylphenols on Hydrocarbon Oxidation Retardation by 4-Methoxyphenol. J. Am. Chem. Soc. 1967, 89-5619—5629.
- Am. Chem. Soc. 1967, 89, 5619-5629.
 (7) Ghaemy, M.; Fruzandeh, S. Synergistic Effects of Some Phosphites Antioxidants Used in Polypropylene Stabilization. Iran Polym. J. 1999, 8, 51-59.
- (8) Xi. F.; Barclay, L. R. C. Cooperative Antioxidant Effects of Ascorbate and Thiols with Di-tert butylcatechol During Inhibited Peroxidation in Solution and in Sodium Dodecyl Sulfate (SDS) Micelles. Can. J. Chem. 1998, 76, 171-182.
 (9) Waterhouse, A. L.; Saucier, C. Antioxidant Synergy Between
- (9) Waterhouse, A. L.; Saucier, C. Antioxidant Synergy Between Phenolics and Sulfites. Book of Abstracts, 218th ACS National Meeting, New Orleans, LA, Aug 22–26, 1999; AN 1999:539298.
- (10) For an analysis of the rate consequences of making intermolecular interactions intramolecular, see: Page, M. I. Energetics of Neighbouring Group Participation. Chem. Soc. Rev. 1973, 2, 295–323.
- (11) Edward, J. T.; Chubb, F. L.; Sangster, J. Iron Chelators of the Isonicotinoyl Hydrazone Class. Relationship of the Lipophilicity of the Apochelator to its Ability to Mobilize Iron From Reticulocytes in Vitro: Reappraisal of Reported Partition Coefficients. Can. J. Physiol. Pharmacol. 1997, 75, 1362-1368.
 (12) Compounds developed specifically to target Fe chelators to
- (12) Compounds developed specifically to target Fe chelators to biological membranes by coupling a chelating moiety to a hydrophobic steroid have been reported; Braughler, J. M.; Burton, P. S.; Chase, R. L.; Pregenzer, J. F.; Jacobsen, E. J.; VanDoornik, F. J.; Tustin, J. M.; Ayer, D. E.; Bundy, G. L. Novel Membrane Localized Iron Chelators as Inhibitors of Iron-Dependent Lipid Peroxidation. *Biochem. Pharmacol.* 1988, 37, 3853-3860.
- (13) Das, N. P.; Ratty, A. K. Studies on the Effect of the Narcotic Alkaloids, Cocaine, Morphine and Codeine on Nonenzymatic Lipid Peroxidation in Rat Brain Mitochondria. Biochem. Med. Metab. Biol. 1987, 37, 256-264.
- (14) (a) Malcolm, C. S.: Benwell, K. R.; Lamb, H.; Bebbington, D.; Porter, R. H. P. Characterization of Iodoacetate Mediated Neurotoxicity In Vitro Using Primary Cultures of Rat Cerebellar Granule Cells. Free Radical Biol. Med. 2000, 28, 102-107. (b) Uto, A.; Dux, E.: Kusumoto, M.; Hossmann, K. A. Delayed Neuronal Death After Brief Histotoxic Hypoxia In Vitro. J. Neurochem. 1995, 65, 2185-2192.
- (15) Kontoghiorghes, G. J.; Lackson, M. J.; Lunec, J. In Vitro Screening of Iron Chelators Using Models of Free Radical Damage. Free Radical Res. Commun. 1986, 2, 115–124.
 (16) Kayyali, R.; Pannala, A. S.; Khodr, H.; Hider, R. C. Comparative
- (16) Kayyali, R.; Pannala, A. S.; Khodr, H.; Hider, R. C. Comparative Radical Scavenging Ability of Bidentate Iron(III) Chelators. *Biochem Pharmacol.* 1998, 55, 1327-1332.
- (17) (a) Kim, D. M.; Kummerow, K. The Antioxidant Activity of 3,5-Di-tert-butyl-4-hydroxybenzyl Derivatives. J. Am. Oil Chem. Soc. 1962, 39, 150–155. (b) Priyadarsini. K. I.; Guha, S. N.; Rao, M. N. A. Physico-Chemical Properties and Antioxidant Activities of Methoxy Phenols. Free Radical Biol. Med. 1998, 24, 933–941.

- (18) The low-yielding insertion reaction of primary amines with 6 is well-reported. For example, see: Rai, B. L.; Dekhordi, L. S.; Khodr, H.; Jin, Y.; Liu, Z.; Hider, R. C. Synthesis. Physicochemical Properties, and Evaluation of N-Substituted-2-alkyl-3hydroxy-4(1H)-pyridinones. J. Med. Chem. 1998, 41, 3347-3359, In our hands, reaction between amines 4 (1 equiv) and benzylmaltol 6 (1 equiv) results in product (24-94%) plus unreacted amine, while f 6 is consumed. Yields were not improved by either slow addition of 6 to a refluxing solution of amine, lower reaction temperatures, changes in solvent, or use of excess 6.
- (19) Dobbin, P. S.; Hider, R. C.; Hall, A. D.; Taylor, P. D.; Sarpong, P.; Poter, J. B.; Xiao, G.; van der Helm, D. Synthesis, Physico chemical Properties, and Biological Evaluation of N-Substituted 2-Alkyl-3-hydroxy-4(1*H*]-pyridinones: Orally Active Iron Chelators with Clinical Potential. *J. Med. Chem.* **1993**, *36*, 2448–2458.
- (20) O'Neill, M. J.; Hicks, C.; Ward, M.; Panetta, J. A. Neuroprotective Effects of the Antioxidant LY231617 and NO Synthase Inhibitors in Global Cerebral Ischemia, Brain Res. 1997, 760, 170 - 178
- (21) Wang, H.; Joseph, J. A. Quantifying Cellular Oxidative Stress by Dichlorofluorescein Assay Using Microplate Reader. Free Radical Biol. Med. 1999, 27, 612-616.
- (a) Whittemore, E. R.; Loo, D. T.; Cotman, C. W. Exposure to Hydrogen Peroxide Induces Cell Death via Apoptosis in Cultured Rat Cortical Neurons. *NeuroReport* **1994**. *5*, 1485–1488. (b) Hoyt, K. R.; Gallagher, A. J.; Hastings, T. G.; Reynolds, I. J.

- Characterisation of Hydrogen Peroxide Toxicity in Cultured Rat
- Forebrain Neurons. Neurochem. Res. 1997, 22, 333-340.
 (23) For example, see: Jarrott, B.; Callaway, J. K.; Jackson, W. R.; Beart, P. M. Development of a Novel Arylalkylpiperazine Compound (AM-36) as a Hybrid Neuroprotective Drug. *Drug* Dev. Res. 1999, 46, 261-267. Ohkawa, S.; Fukatsu, K.; Miki, S.; Hashimoto, T.; Sakamoto, J.; Doi, T.; Nagai, Y.; Aono, T. 5-Aminocoumarins: Dual Inhibitors of Lipid Peroxidation and Dopamine Release with Protective Effects against Central Nervous System Trauma and Ischemia. J. Med. Chem. 1997, 40, 559-573. Chabrier, P.-E.; Auguet, A.; Spinnewyn, B.; Auvin, S.; Cornet, S.; Demerle Pallardy, C.; Guilmard-Favre, C.; Marin, J.-G.; Pignol, B.; GillardRoubert, V.; Roussillot-Charnet, C.; Schulz, J.; Viossat, I.; Bigg, D.; Moncada, S. BN 80933, A Dual Inhibitor of Neuronal Nitric Oxide Synthase and Lipid Peroxidation: A Promising Neuroprotective Strategy. Proc. Natl. Acad. Sci. U.S.A. 1999, 96, 10824–10829.
- (24) The biological activity of the protected 3-hydroxy-4(1H)-pyridinone 10 (which theoretically has no potential to chelate Fe) indirectly provides an indication of the likely individual contributions afforded by the two structural units of 3b to the inhibition of lipid peroxidation and cell toxicity (compd 10: LP IC_{50} 4.5 μ M; IAA cell toxicity EC_{50} 12.8 μ M (relative efficacy 0.5)).
- This work is the subject of International Patent Number WO99/23075.

JM990945V